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FORM PTO-1390 (REV. 11-2000) U S DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371

ATTORNEY 'S DOCKET NUMBER: HERR 18.313

U S APPLICATION NO. (If known, see 37 CFR 1 5): 09/762283

INTERNATIONAL APPLICATION NO.	INTERNATIONAL FILING DATE	PRIORITY DATE CLAIMED
PCT/ES00/00197	02 JUNE 2000 (02.06.00)	04 JUNE 1999 (04.06.99)

TITLE OF INVENTION
METHOD FOR TREATING ENDOTOXIC SHOCK AND INFLAMMATORY AND AUTOIMMUNE DISEASES IN MAMMALS

APPLICANT(S) FOR DO/EO/US
Rosa PEREZ GOMARIZ, et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. [X] This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. [ ] This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. [X] This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.
4. [ ] The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. [X] A copy of the International Application as filed (35 U.S.C. 371(c)(2))
   a. [ ] is attached hereto (required only if not communicated by the International Bureau).
   b. [X] has been communicated by the International Bureau.
   c. [ ] is not required, as the application was filed in the United States Receiving Office (RO/US).
6. [ ] An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
   a. [ ] is attached hereto.
   b. [ ] has been previously submitted under 35 U.S.C. 154(d)(4).
7. [X] Amendments to the claims of the International Aplication under PCT Article 19 (35 U.S.C. 371(c)(3))
   a. [ ] are attached hereto (required only if not communicated by the International Bureau).
   b. [ ] have been communicated by the International Bureau.
   c. [ ] have not been made; however, the time limit for making such amendments has NOT expired.
   d. [X] have not been made and will not be made.
8. [ ] An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371 (c)(3)).
9. [ ] An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. [ ] An English lanugage translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 20 below concern document(s) or information included:

11. [ ] An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. [ ] An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. [ ] A FIRST preliminary amendment.
14. [ ] A SECOND or SUBSEQUENT preliminary amendment.
15. [ ] A substitute specification.
16. [ ] A change of power of attorney and/or address letter.
17. [ ] A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
18. [ ] A second copy of the published international application under 35 U.S.C. 154(d)(4).
19. [ ] A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
20. [ ] Other items or information:

Any fee due with this paper, not fully covered by an enclosed check, may be charged on Deposit Acct. No. 08-1634.

Filed by Express Mail (Receipt No. EL522399274US) on February 5, 2001 pursuant to 37 C.F.R. 1.10. by

JC03 Rec'd PCT/PTO 05 FEB 2001

U.S. APPLICATION NO (if known, see 37 CFR 1.5)	INTERNATIONAL APPLICATION NO	ATTORNEY'S DOCKET NUMBER
09/762283	PCT/ES00/00197	HERR 18.313

21. [X] The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)):		CALCULATIONS PTO USE ONLY
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO	$1000.00	
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO	$860.00	
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO	$710.00	
International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4)	$690.00	
International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4)	$100.00	
ENTER APPROPRIATE BASIC FEE AMOUNT =		$ 1,000.00
Surcharge of **$130.00** for furnishing the oath or declaration later than [X] 20 [ ] 30 months from the earliest claimed priority date (37 CFR 1.492(e)).		$ 130.00

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	$
Total claims	9 - 20 =	0	x **$18.00**	$ 0.00
Independent claims	3 - 3 =	0	x **$80.00**	$ 0.00
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ **$270.00**	$
TOTAL OF ABOVE CALCULATIONS =				$ 1,130.00
[ ] Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.			+	$
SUBTOTAL =				$ 1,130.00
Processing fee of **$130.00** for furnishing the English translation later than [X] 20 [ ] 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				$ 130.00
TOTAL NATIONAL FEE =				$ 1,260.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). **$40.00** per property +				$
TOTAL FEES ENCLOSED =				$ 1,260.00
			Amount to be refunded:	$
			charged:	$

a. [X] A check in the amount of $ 1,260.00 to cover the above fees is enclosed.

b. [ ] Please charge my Deposit Account No. _____ in the amount of $ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. [X] The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 08-1634 . A duplicate copy of this sheet is enclosed.

d. [ ] Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137 (a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

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23,072

REGISTRATION NUMBER

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On: March 28, 2001
By Doreen DeWitt
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Attorney Docket No.: HERR 18.313

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Inventor	:	**Rosa PEREZ GOMARIZ, et al.**
Serial No.	:	**09/762,283**
Filed	:	**February 5, 2001**
Title	:	**METHOD FOR TREATING ENDOTOXIC SHOCK AND INFLAMMATORY AND AUTOIMMUNE DISEASES IN MAMMALS**

March 28, 2001

Assistant Commissioner for Patents
Washington, D.C. 20231

PRELIMINARY AMENDMENT

S I R:

Prior to the issuance of an Office Action, amend the application as follows:

In the Claims

Please add the following claims:

--10. Use of the vasoactive intestinal peptide (VIP) or any of its fragments or some analogue derivative for the preparation of a drug destined for the treatment of endotoxic shock in mammals, due to their capacity as agents that inhibit the production of the tumoral necrosis factor (TNF) and interleukin 6 (IL-6).

11. Use of the adenylate cyclase hypophysary activator peptide (ACHPA) or a fragment thereof or a derivative for the preparation of a drug destined to the treatment of endotoxic shock in mammals, due to their capacity as agents that inhibit the production of the tumoral necrosis factor (TNF) and interleukin 6 (IL-6).

The Commissioner is hereby authorized to charge any additional fees under 37 CFR 1.16 and 1.17 which may be required during the entire pendency of the application to Deposit Account No 08-1634. EXCEPT THE ISSUE FEE.

12. Use of the vasoactive intestinal peptide (VIP) or any of its fragments or some analogue derivative for the preparation of a drug destined to the treatment of inflammatory or autoimmune pathologies characterized by the activation of Th1 cells, such as rheumatoid arthritis, multiple sclerosis, Crohn's disease, implant reaction to host and others, due to their capacity as inhibitors of Th1 cells.

13. Use of the adenylate cyclase hypophysary activator peptide (ACHPA) or a fragment thereof or a derivative for the preparation of a drug destined to the treatment of inflammatory or autoimmune pathologies characterized by the activation of Th1 cells, such as rheumatoid arthritis, multiple sclerosis, Crohn's disease, implant reaction to host and others, due to their capacity as inhibitors of Th1 cells. --

REMARKS/ARGUMENTS

Claims 1-9 remain in this application. Claims 10-13 were added.

Any fee due with this paper, not fully covered by an enclosed check, may be charged on Deposit Account 08-1634.

Respectfully submitted,

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COMPOSITION AND METHOD FOR THE TREATMENT OF ENDOTOXIC SHOCK AND INFLAMMATORY AND AUTOIMMUNE DISEASES IN MAMMALS.

STATE OF THE ART

Endotoxic shock is still the main cause of death in hospitals. The strategies for combating the effects of endotoxic shock focus on counteracting the bacterial agents responsible for the effect, on restoring the haemodynamic parameters, preventing cellular activation and on modifying the action of the defence mechanisms (Boyd O; Current Opinion in Anaesthesiology 1996, 9:98).

It is currently accepted that the inflammatory response to bacterial products directly contributes to the development of endotoxic shock (Parillo JE; New England Journal of Medicine 1993, 328:1471). The toxic bacterial products and those released during tissue damage activate the defence mechanisms, implicating cells such as neutrophils, monocytes, macrophages and endothelial cells, and mediators such as cytokines, platelet activation factor, metabolites of arachidonic acid and nitric oxide, causing haemodynamic changes and organics harmful to the host (Moldawer LL; Critical Care Medicine 1994, 22:3). Many cytokines have been proposed as markers of the seriousness of the development of septic shock. The circulating levels of TNFα, IL-l, IL-6 and IL-8 have been correlated with the probability of overcoming a septic episode. TNFα and IL-1 administered to humans or to experimental animals reproduce many of the haemodynamic manifestations of septic shock (Tracey KJ and co-workers; Science 1986, 234:470) and its inhibition has been assayed by means of injection of antagonist receptors and neutralising monoclonal antibodies with different results (Fisher CJ and co-workers; Critical Care Medicine 1994, 22:12). Among the immunological markers, the levels of circulating IL-6 are the best indicators of the seriousness of the sepsis and

of the possibilities of recovering from the episode (Liaw YS and co-workers.; Journal of the Formosan Medical Association 1997, 96:685).

Despite the advance in the knowledge of the mechanisms and of the technological and pharmacological progress there are still few results concerning an improvement in the data for death, which translate into a figure of around 200,000 per year in the United States and Europe (Vicent J-L and Chamlou R; Current Opinion in Anaesthesiology 1996, 9:146).

Inflammatory processes are a vital process for the survival of all complex organisms. Inflammation is a natural defence process of the organism against foreign agents. The accumulation and activation of leukocytes in places where the aggression takes place is a central occurrence in all inflammatory processes (Schaal TJ and Bacon KB; Current Opinion in Immunology 1994, 6:865). An insufficient inflammatory response may compromise the survival of the organism, but an excessive response, which may be due to failures in the mechanisms of deactivation of the process due to different causes, can lead to an inflammatory or autoimmune disease (Sacca R and co-workers; Current Opinion in Immunology 1997, 9:851). These diseases are an important cause of morbidity and mortality in mammals due to tissue damage associated with said processes.

Macrophages play a key role in the regulation of immune and inflammatory responses. The execution of these activities is mediated by a whole series of complex processes in which, among others, many products of macrophage origin intervene. As a response to the antigens, and according to their origin, the macrophages secrete pro-inflammatory cytokines and oxidising agents, such as TNFα, IL-6, IL-1β, IL-12 and nitric oxide (Laskin DL and co-workers; Annual Review of Pharmacology and Toxicology 1995, 35:655). TNFα and IL-6 are, among others, two factors that contribute to physiopathological changes associated with several states of chronic or acute

inflammation. The macrophages, in addition, participate in the initiation, maintenance and control of immune responses, acting as potent antigen presenters, providing the T-lymphocytes with a double activation signal: the antigen-molecule complex of the main histocompatibility complex (MHC) and a co-stimulatory signal mediated by molecules of the B7 family (Lenschow DJ and co-workers; Annual Review of Immunology 1996, 14:233). The B7 molecules comprise two isoforms, B7.1 and B7.2, both of which are implicated in the stimulation of two different types of T helper cells (Th), Th1 and Th2, and both of these produces a set of different cytokines (Kuchroo VK and co-workers; Cell 1995, 80:707).

Activation of Th1 cells implies the production of IFNγ and IL-12, among other factors, and is associated with the production of antibodies of the IgG2a isotype and it is manifest as a reaction of delayed inflammatory type. The activation of Th2 cells implies the production of IL-4, IL-5 and IL-10, among other factors, is associated with the secretion of antibodies of the isotype IgG1, inhibits the delayed inflammatory response and is manifest as a humoral response (Constant SL y Bottomly K; Annual Review of Immunology 1997, 15:297). The factors that determine the differentiation of one or an other type of response are mainly the characteristics of the antigen presenting cells and the cytokines present in the microenvironment in which the response takes place: IL-12 determines the differentiation of Th1 cells while IL-4 does so with Th2. When both are present, the IL-4 effect predominates (O'Garra AO; Immunity 1998, 8:275). Numerous cases of inflammatory and autoimmune diseases are due to the activation of an inappropriate type of Th cell.

The vasoactive intestinal peptide (VIP) is a basic peptide containing 28 amino acids units whose sequence is (Mutt V and Said SI; European Biochemistry 1974, 42:581):

His-Ser-Asp-Ala-Val-Phe-Thr-Asp-Asn-Tyr-Thr-Arg-Leu-Arg-Lys-Gln-Met-Ala-Val-Lys-Lys-Tyr-Leu-Asn-Ser-Ile-Leu-Asn-NH2

It was isolated firstly from the small intestine of pig and later was identified in the brain and in the nerve endings of the peripheral system. It was established that it was a neuropeptide with neuromodulating properties (Fahrenkrug J; Pharmacology and Toxicology 1993, 72:354). It takes its name from its peripheral vasodilatory properties. VIP has also been identified in mast cells of rat and in granulomas (Cutz E. and co-worders: Nature 1978, 275:661). Immunochemical studies carried out in histological sections of the thymus, spleen and lymphatic ganglia of rat have identified immunoreactive VIP in lymphocytes of these organs (Gomariz RP and co-workders; Annals of the New York Academy of Sciences 1992, 650:13; Leceta and co-workers; Advances in Neuroimmunology 1996, 6:29).

VIP exercises its biological effects by means of membrane receptors belonging to the superfamily of seven hydrophobic domains coupled to G proteins, which transduce the information to the final effecter molecules (Laburthe M and Couvineau A; Annals of the New York Academy of Sciences 1988, 527:296). The receptors for VIP have been characterised in numerous tissues such as liver and adipose tissue, among others, and correspond to two types, the so-called VIP1 -R (Ishihara T and co-workers; Neuron 1992, 8:811) and VIP2-R (Lutz E. and co-workers; FEBS Letters 1993, 334:3). In the immune system, receptors specific to VIP have been characterised in a variety of immune cells which include human peripheral lymphocytes, human monocytes, rat and mouse lymphocytes, alveolar macrophages of rat and peritoneal macrophages of rat and mouse (Gomariz RP and co-workers; Biochemical and Biophysical Research Communications 1994, 203:1599; Delgado M and co-workers; Regulatory Peptides 1996, 62:161). VIP modulates a wide variety of immune functions such as the phagocyte function, in each one of the stages of the process, the proliferate response, the production of immunoglobins, the NK activity and the production of cytokines (De La Fuente M and co-workers; Advances in Neuroimmunology 1996, 6:75).

The adenylate cyclase hypophysary peptide activator (ACHPA) is a member of the family of peptides of the secretin/VIP/glucagons of which two molecular forms are known: ACHPA-38 and ACHPA-27, whose sequences are respectively (Ogi K and co-workers: Biochemical and Biophysical Research communication 1993, 196:1511):

ACHPA-38

His-Ser-Asp-Gly-Ile-Phe-Thr-Asp-Ser-Tyr-Ser-Arg-Tyr-Arg-Lys-Gln-Met-Ala-Val-Lys-Lys-Thy-Leu-Ala-Ala-Val-Leu-Gly-Lys-Arg-Tyr-Lys-Gln-Arg-Val-Lys-Asn-Lys-NH2

ACHPA-27

His-Ser-Asp-Gly-Ile-Phe-Thr-Asp-Ser-Tyr-Ser-Arg-Tyr-Arg-Lys-Gln-Met-Ala-Val-Lys-Lys-Thy-Leu-Ala-Ala-Val-Leu-NH2

Both peptides are widely distributed in the central nervous system and peripheral system.

There are also cells that produce ACHPA in the lung, pancreatic B cells and intestine (Arimura A: Regulatory Peptides 1992, 37:287). In the immune system, a large abundance of ACHPA positive cells has been described in central and peripheral lymphoid organs (Gaytan F and co-workers; Cell and Tissue Research 1994, 276:233). For ACHPA, three types of receptor have been described (Shivers BD and co-workers; Endocrinology 991, 128:3055; Inagaki N and co-workers; Proceeding of the National Academy of Sciences USA 1994, 91:2679). The type-1 ACHPA (ACHPA-R-I) with equal affinity for ACHPA-38 and ACHPA-27, but which has an affinity of 300 to 1000 times less for VIP; the type-2 ACHPA receptor (ACHPA-R-II) which recognises VIP, ACHPA-38 and ACHPA-27 with the same affinity, and so is denominated the VIP-ACHPA common receptor and corresponds to the receptor of VIP VIPl-R, and the type-

III ACHPA receptor (ACHPA-R-III) which corresponds to the receptor of VIP VIP2-R. Until present, there are few studies on the biological actions of ACHPA in the immune system. The effects of ACHPA are often similar to those of VIP modulating the phagocyte function and the proliferate responses.

DESCRIPTION OF THE INVENTION

The object of this invention is to develop preparations of VIP, ACHPA and analogues as therapeutic agents in the treatment of endotoxic shock and inflammatory and autoimmune diseases.

The treatment consists of the administration to mammals, in need thereof, of an effective quantity of an agent that inhibits the production of tumoral necrosis factor (TNF) or IL-6 in a pharmaceutically acceptable vehicle, or else the administration to mammals, in need thereof, of an effective quantity of an agent that increases the production of IL-4, inhibiting the activation of Th1 cells and stimulating the activation of Th2 cells.

It is known that most of the effects of endotoxic shock are mediated by the activation of the immune system and the inflammatory mechanisms of the host as a response to the bacterial products. The macrophages play a very relevant role in this process as, after their activation, they produce factors such as nitric oxide, prostaglandins and cytokines which are responsible for symptoms such as fever, hypotension, disseminated microcoagulation, multiple organ failure and finally death. In this sense, high circulating levels of TNF, IL-1 and IL-6 associated with endotoxaemia have been described. In animal models these symptoms are reproduced both by the administration of bacterial endotoxins (LPS) and by injection of TNF and IL-1. Other studies have shown the diagnostic value in terms of probability of survival that circulating levels of IL-6 represent (Stoiser B and co-workers; European Journal of Clinical Investigation 1998, 28:672).

The tumoral necrosis factor (TNF) is produced by several types of cell which include monocytes and macrophages, T and B lymphocytes, neutrophils, mast cells, tumoral cells and fibroblasts. It is an important regulatory factor of other pro-inflammatory cytokines, such as IL-1β, IL-6 and IL-8. TNFα induces the expression of adhesion molecules in endothelial cells, activates the leukocytes so that they destroy the microorganisms, acts on the hepatocytes in order to increase synthesis of serum proteins which contribute to the response in acute phase and activate the coagulation system. Overproduction of this molecule leads to immunopathological diseases, autoimmunity and inflammation.

IL-6 is a multifunctional cytokine produced both by lymphocytes and by non-lymphoid cells. It regulates several aspects of the immune response, such as production of proteins that mediate the acute phase of haematopoiesis. In addition, it acts as a mediator in the inflammatory response. Its production is regulated by several factors which include TNFα, IL-1 and bacterial endotoxin (LPS).

IL-4 is a cytokine that inhibits the production of pro-inflammatory cytokines, promotes the proliferation and differentiation of activated lymphocyte B molecules and increases the expression of MHC molecules of type II and B lymphocytes. Its possible clinical use in anti-inflammatory treatment and autoimmune diseases has been highlighted.

Strategies have been tried for neutralising pro-inflammatory cytokines in the treatment of endotoxic shock but the results do not indicate that there is greater survival in the long term. A treatment that inhibits the production of TNFα and IL-6 would represent a considerable improvement in the evolution of endotoxic shock and in the probabilities of survival. The administration of VIP and ACHPA in animal models achieved these effects and our invention consists in the use of a treatment with these neuropeptides to

increase the survival in endotoxic shock and revert pathological inflammatory states and autoimmune diseases.

VIP and ACHPA have anti-inflammatory effects and inhibit the production of IL-6 and TNFα in animal models of induction of endotoxic shock. As these cytokines play an important role in the development of said syndrome, VIP and ACHPA can be used to regulate their production. In addition, VIP and ACHPA modulate the capacity of the antigen presenting cells to act inducing the activation of proliferation and differentiation of lymphocytes with a pattern of cytokine secretion typical of Th2 cells and condition the immune responses "in vivo" favouring the development of response of the humoral type.

DESCRIPTION OF THE FIGURES

Figure 1 represents the production of TNFα by murine macrophages in culture ($5x10^5$ cells/ml) stimulated with 10ngr/ml of LPS in the presence or absence of 10^{-8}M of VIP or ACHPA during the course of 24 hours.

Figure 2 represents the production of TNFα by murine macrophages in culture ($5x10^5$ cells/ml) after 6 hours of culture with 10ngr/ml of LPS and to which 10^{-8}M of VIP or ACHPA are added at different times.

Figure 3 represents the production of IL-6 by murine macrophages in culture ($5x10^5$ cells/ml) stimulated with 10ngr/ml of LPS in the presence or absence of 10^{-8}M of VIP or ACHPA during the course of 24 hours.

Figure 4 represents the production of IL-6 by murine macrophages in culture ($5x10^5$ cells/ml) after 6 hours of culture with 10ngr/ml of LPS and to which 10^{-8}M of VIP or ACHPA are added at different times.

Figure 5 presents the Northern Blot analysis for the presence of mRNA of TNFα and IL-6 in macrophages stimulated with LPS in the presence or absence of VIP or ACHPA (18S represents the corresponding rRNA as total quality control of loaded RNA).

Figure 6 represents the survival in mice injected with 400μgr of LPS and, simultaneously, or after 30 minutes, 1 or 4 hours, with 5 nmol of VIP or ACHPA.
A. Control; B: VIP at 0h.; C: VIP at 0,5 h; D: VIP at 1 h.; E: VIP at 4 h.

Figure 7 represents the number of IL-4 secreting cells in the spleen and peritoneum detected by means of the immunabsorption assay technique in conjugated plate with enzymes (ELISPOT) in mice immunised in the conditions specified in Example 7 and which simultaneously to the second injection of antigen received 5 nmol of VIP or ACHPA or an injection of saline solution.

Figure 8 represents the quantity of anti-haemocyanine immunoglobulins of snail (anti-KLH) of the isotypes IgG2a and IgG1 detectable in serum by means of the technique conjugated immunoabsorption assays with enzymes (ELISA) in immunised mice in the conditions specified in Example 8 and with the serum samples taken two weeks after the last injection.

Figure 9 represents the number of IL-4 producing cells detected by means of the ELISPOT technique in mice that were immunised in the conditions specified in Examples 7 and 8 and which, in the second injection, received or did not receive 5 nmol of VIP along with 100μgr of IgG, anti-B7.1 or anti-B7.2.

EMBODIMENT OF THE INVENTION

The following examples are only to illustrate the results obtained and do not limit the use of the invention which is detailed in the specified claims.

EXAMPLE 1

VIP and ACHPA inhibit the production of TNFα in macrophages stimulated with LPS

In experiments carried out "in vitro" VIP and ACHPA inhibit the production of TNFα in peritoneal murine macrophages stimulated with LPS. The greatest degree of inhibition reaches values near to 60% and occurs with doses of stimulation between 1-10 ngr./ml of LPS. The IC_{50} value is 80 pM, both for VIP and for ACHPA and its effect was observed up until the end of the experiment (see Figure 1). The inhibitory effect is the same if both neuropeptides are added up until 1 hour after stimulating the macrophages with LPS, although it progressively reduces until disappearing if they are added after 4 hours (see Figure 2).

EXAMPLE 2

VIP and ACHPA reduce the circulating levels of TNFα after injection of LPS

In an experiment carried out with mice the circulating levels of TNFα 2 hours after injection of 25 μgr of LPS were approximately 4 ngr./ml. Simultaneous administration of 5 nmol of VIP or ACHPA reduced said levels by 60%.

EXAMPLE 3

VIP and ACHPA inhibit production of IL-6 in macrophages stimulated with LPS

In experiments carried out "in vitro" VIP and ACHPA inhibit the production of IL-6 in peritoneal murine macrophages stimulated with LPS. Most of the inhibition reaches values near to 90% and occurs with doses of stimulation of 10 μgr./ml of LPS. The IC_{50} is 8.6 pM, both for VIP and for ACHPA and the effect was observed up until the end of

the experiment (see Figure 3). The inhibitory effect is also observed if the neuropeptides are added after stimulation with LPS, although the degree of inhibition is progressively smaller (see Figure 4).

EXAMPLE 4

VIP and ACHPA reduce the circulating levels of IL-6 after injection of LPS

In an experiment carried out in mice the circulating levels of IL-6 two hours after injection of 25 μgr. of LPS were approximately 1.5 ngr./ml. The simultaneous administration of 5nmol of VIP or ACHPA reduced said levels by 60% and 75%, respectively.

EXAMPLE 5

VIP and ACHPA regulate the production of TNFα and IL-6 at a transcriptional level

Macrophages from rat were submitted to the experimental conditions described in examples 1 and 3 and their mRNA was isolated. This mRNA was analysed using the Northern Blot technique to detect mRNA of TNFα and IL-6. Figure 5 shows the absence of transcripts for TNFα or IL-6 when the macrophages activated with LPS are also exposed to VIP or ACHPA.

EXAMPLE 6

VIP and ACHPA protect against the lethal effects of LPS

An experiment was carried out in which the long-term survival over a period of 4 days was studied for mice injected with 400μgr of LPS. The results are reflected in figure 6.

The mortality in these circumstances was 100% after 36 hours. With the simultaneous administration of 5 nmol of VIP or ACHPA a survival of 60% was achieved at the end of the experiment. The administration of neuropeptides up to 1 hour after injection with LPS still gave survival rates close to 50%.

EXAMPLE 7

VIP and ACHPA increase the proportion of IL-4 secreting cells.

Groups of mice were immunised with 50 μgr of KLH emulsified in an adjuvant, repeating the injection with 100 μgr of KLH two weeks later and simultaneously injecting 5 nmol/mouse of VIP, ACHPA or saline solution. Two weeks after the last injection, suspensions were made of the spleen and peritoneum cells. These were cultured for 24 hours in the presence of 50 μgr/ml of KLH, after which time the number of IL-4 producing cells was determined using the ELISPOT technique. In the mice injected with VIP or ACHPA, the number of IL-4 producing cells increased in the order of 20 times compared to those that were not treated with these neuropeptides (see Figure 7).

EXAMPLE 8

VIP and ACHPA induce the production of antibodies of the isotype IgG1.

Groups of mice were immunised with 50 μgr of KLH emulsified in an adjuvant, repeating the injection with 100 μgr of KLH two weeks later and simultaneously injecting 5 nmol/mouse of VIP, ACHPA or saline solution. Two weeks after the last injection, the levels of anti-KLH and its isotype were determined using ELISA specific to the IgG1 and IgG2a isotypes. In mice injected with VIP or ACHPA the anti-KLH antibodies detectable in serum two weeks after the last immunisation are only of the

IgG1 isotype. while the in those that only received saline solution they were of the isotype IgG2a (see Figure 8)

EXAMPLE 9

The increase in the proportion of IL-4 producing cells mediated by VIP and ACHPA is related to the expression of B7.2 induced by both neuropeptides.

Groups of mice were immunised in the sample conditions as those for Examples 7 and 8, but in the moment of the second immunisation with KLH the mice that were simultaneously injected with VIP or ACHPA also received 100 μgr of anti-B7.1, anti-B7.2 antibody or the same quantity of IgG as control. In the mice that received anti-B7.2 antibodies simultaneously to the administration of neuropeptides the number of IL-4 producing cells was reduced to the proportion reached in animals that were not injected with neuropeptides (see Figure 9).

CLAIMS

1.- A method for the treatment of endotoxic shock in mammals characterised in that it comprises the administration of an effective quantity of an agent that inhibits the production of tumoral necrosis factor (TNF) in a pharmaceutically acceptable vehicle.

2.- A method for the treatment of endotoxic shock in mammals according to claim 1, characterised in that the inhibitory agent is a vasoactive intestinal peptide (VIP) or any fragments thereof or some analogue derivative.

3.- A method for the treatment of endotoxic shock in mammals according to claim 1, characterised in that the inhibitory agent is the adenylate cyclase hypophysary peptide activator (ACHPA) or any fragments thereof or some analogue derivative.

4.- A method for the treatment of endotoxic shock in mammals characterised in that it comprises the administration of an effective quantity of an agent that inhibits the production of interleukin 6 (IL-6) in a pharmaceutically acceptable vehicle.

5.- A method for the treatment of endotoxic shock in mammals according to claim 4, characterised in that the inhibitory agent is a vasoactive intestinal peptide (VIP) or any fragments thereof or some analogue derivative.

6.- A method for the treatment of endotoxic shock in mammals according to claim 4, characterised in that the inhibitory agent is the adenylate cyclase hypophysary peptide activator (ACHPA) or any fragments thereof or some analogue derivative.

7.- A method for the treatment of inflammatory or autoimmune pathologies in mammals, characterised by the activation of Th1 cells, which comprises the administration of an

effective dose of an agent, in a pharmaceutically appropriate vehicle, which induces high levels of IL-4.

8.- A method for the treatment of inflammatory or autoimmune pathologies in mammals according to claim 7, characterised in that the inducing agent is the vasoactive intestinal peptide (VIP) or a fragment thereof or some analogue derivative.

9.- A method for the treatment of inflammatory or autoimmune pathologies in mammals according to claim 7, characterised in that the inducing agent is the adenylate cyclase hypophysary activator peptide (ACHPA) or any fragment thereof or some analogue derivative.

ABSTRACT

The composition and use of therapeutic agents that inhibit the production of tumoral necrosis factor and interleukin 6 are described. Said agents induce high levels of interleukin 4 for the treatment of endotoxic shock and inflammatory and autoimmune diseases, respectively, in mammals. The composition includes substances such as vasoactive intestinal peptide, adenylate cyclase hypophysary activator peptide and fragments and derivatives thereof.

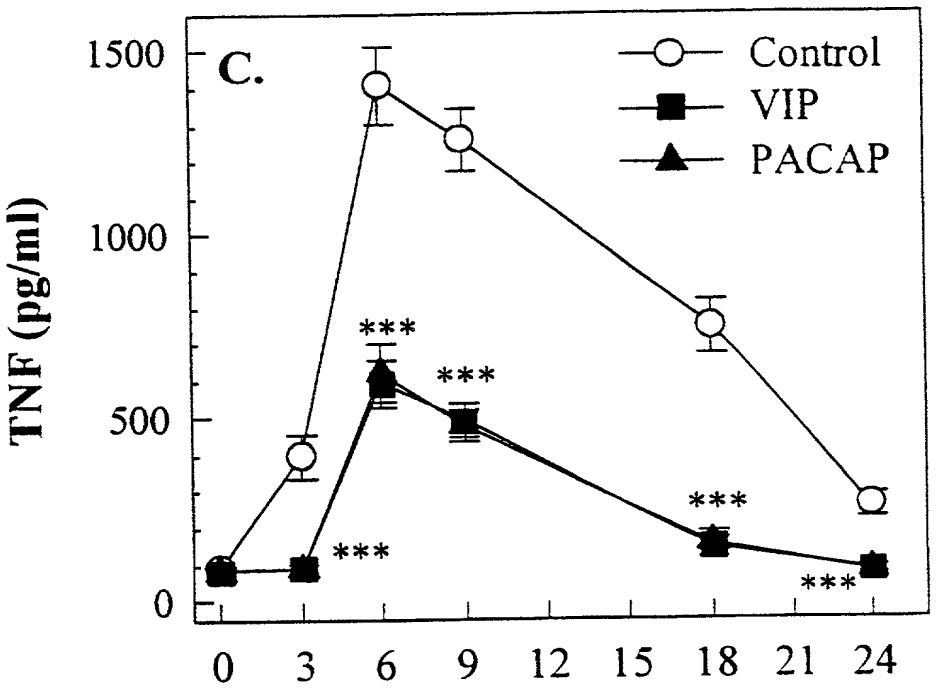

FIGURE 1

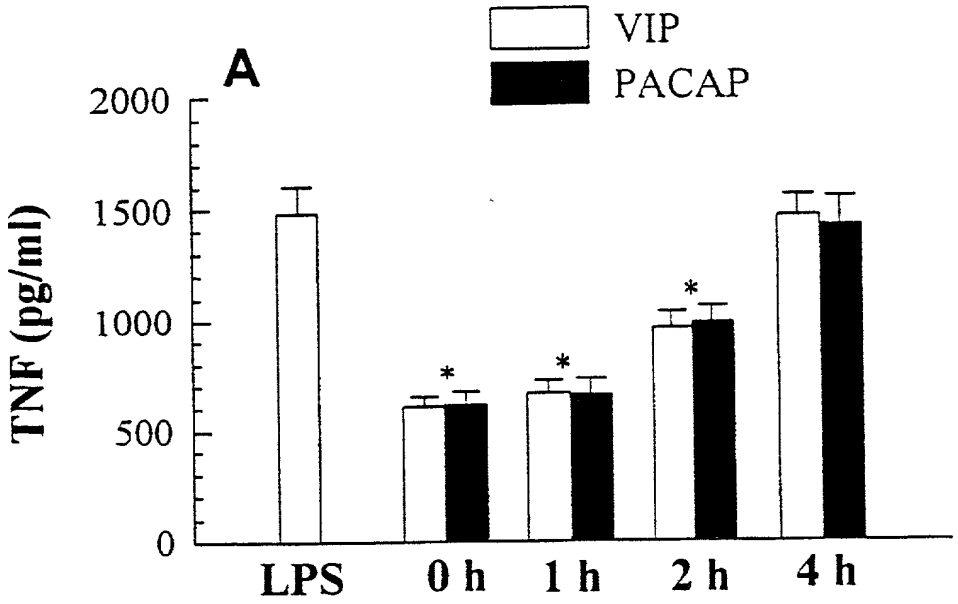

FIGURE 2

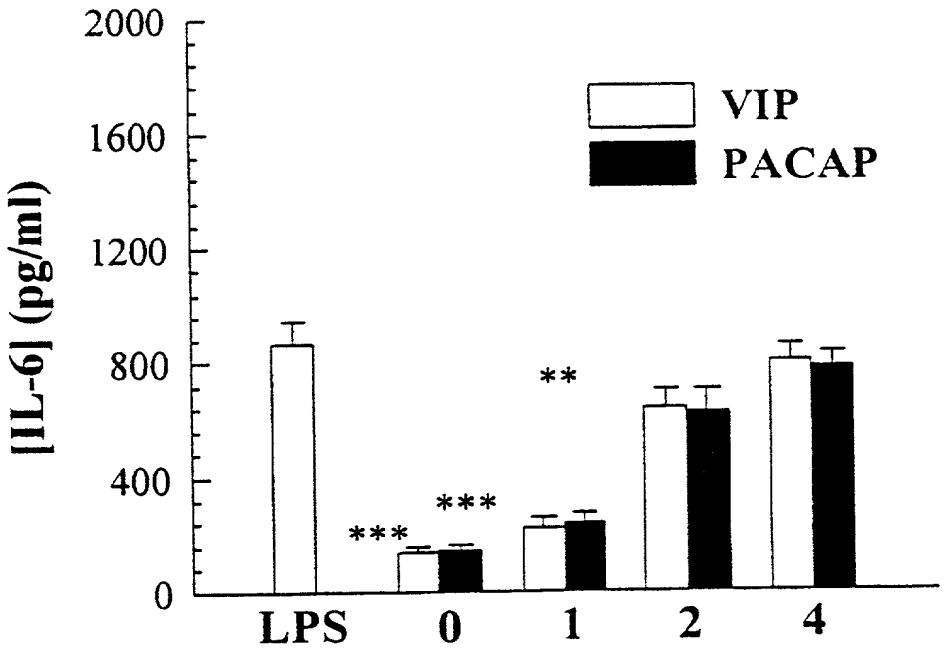

FIGURE 3

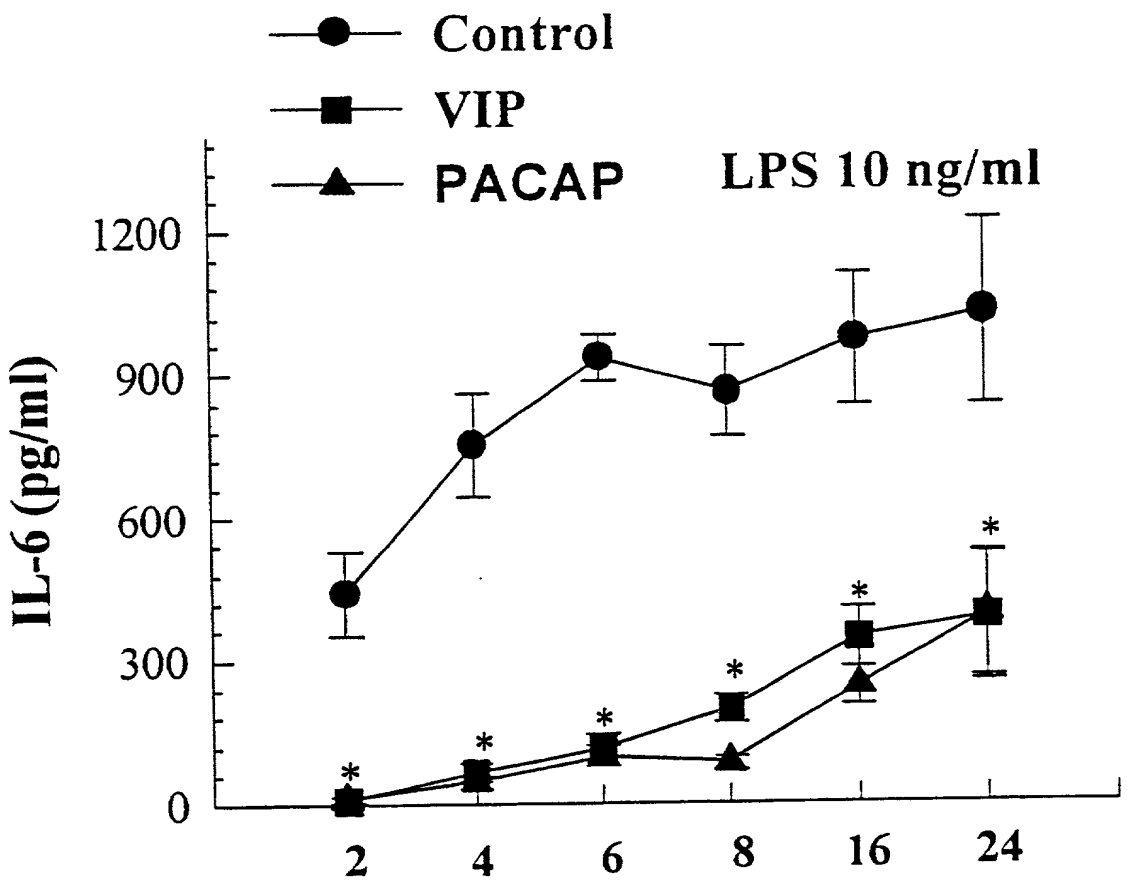
Control
VIP
PACAP
LPS 10 ng/ml
IL-6 (pg/ml)
1200
900
600
300
0
2
4
6
8
16
24
*

FIGURE 4

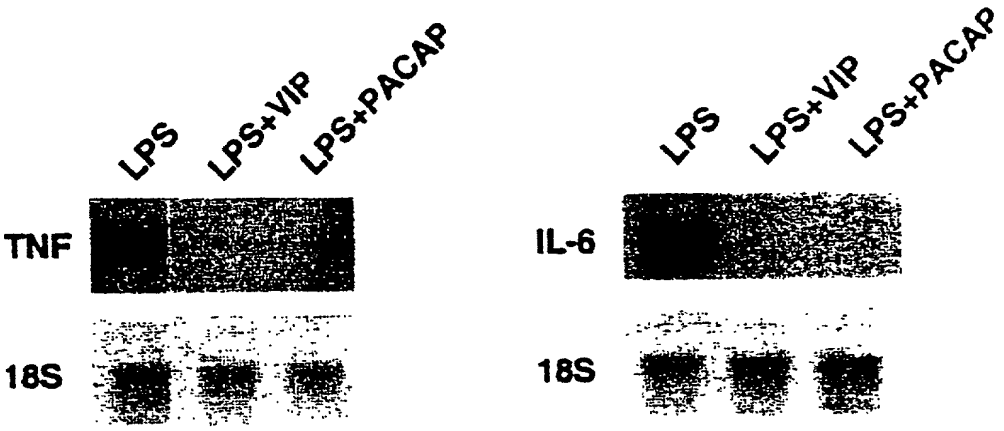

FIGURE 5

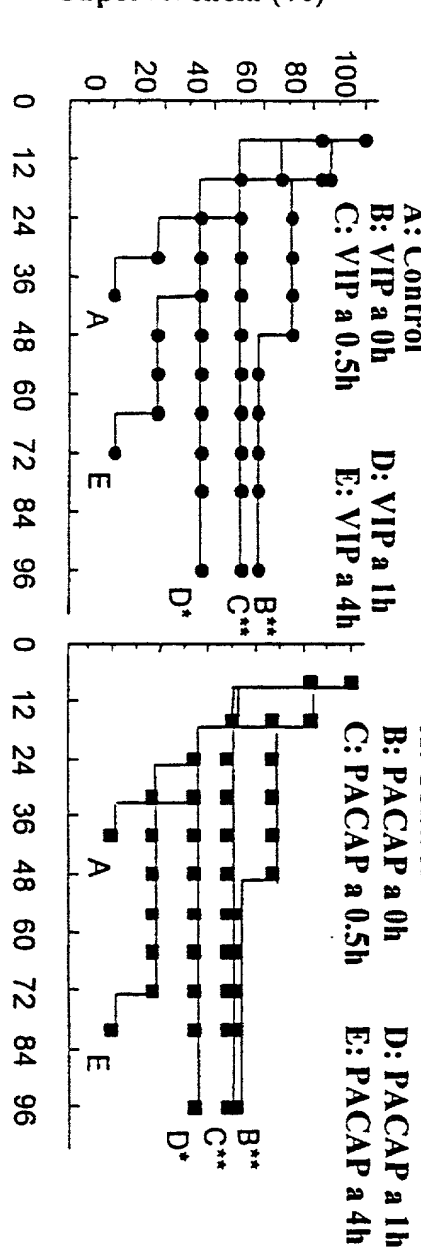

FIGURE 6

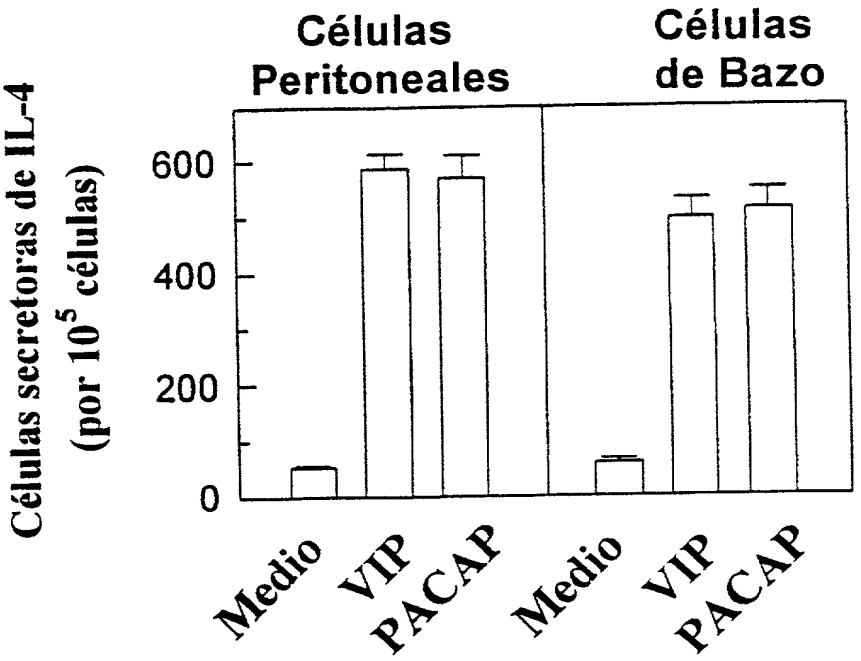
Células Peritoneales
Células de Bazo
Células secretoras de IL-4 (por 10^5 células)
600
400
200
0
Medio
VIP
PACAP
Medio
VIP
PACAP

FIGURE 7

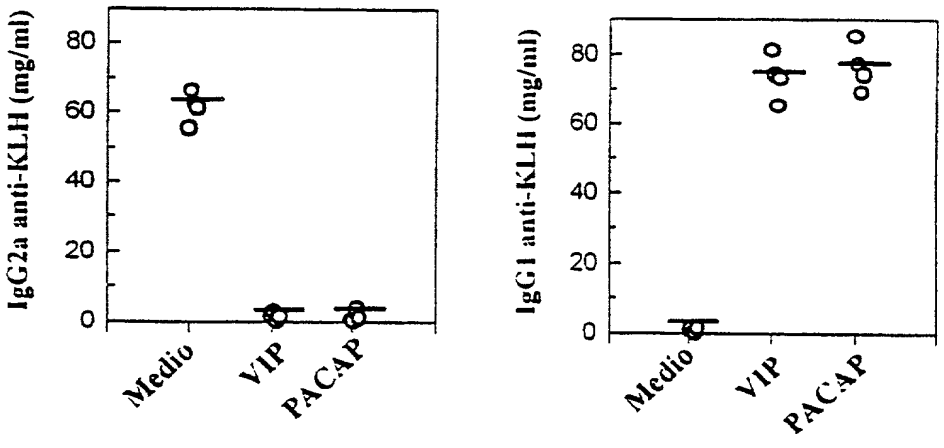

FIGURE 8

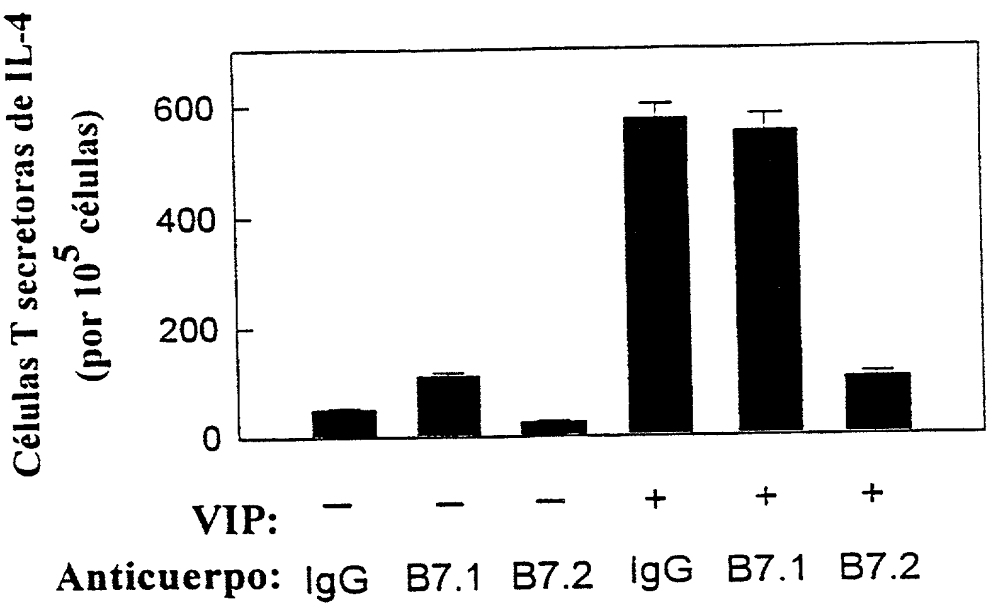
Células T secretoras de IL-4
(por 10^5 células)
600
400
200
0
VIP:
−
−
−
+
+
+
Anticuerpo:
IgG
B7.1
B7.2
IgG
B7.1
B7.2

FIGURE 9

Combined Declaration for Patent Application and Power of Attorney

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

COMPOSITION AND METHOD FOR THE TREATMENT OF ENDOTOXIC SHOCK AND INFLAMMATORY AND AUTOIMMUNE DISEASES IN MAMMALS

, the specification of which

(check one)		
		is attached hereto.
	X	was filed on Feb. 5, 2001 as Application Serial No. 09/762,283 and (if applicable) was amended on:

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed	
9901235 (Number)	SPAIN (Country)	04 June 1999 (Day Month Year Filed)	X	
(Number)	(Country)	(Day Month Year Filed)	YES	NO
(Number)	(Country)	(Day Month Year Filed)	YES	NO
(Number)	(Country)	(Day Month Year Filed)	YES	NO
(Number)	(Country)	(Day Month Year Filed)	YES	NO
(Number)	(Country)	(Day Month Year Filed)	YES	NO
(Number)	(Country)	(Day Month Year Filed)	YES	NO

I hereby claim the benefit under Title 35, United States Code, §120 of any United States Application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulation, §1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application.

(Application Serial No.)	(Filing Date)	(Status: patented, pending, abandoned)
(Application Serial No.)	(Filing Date)	(Status: patented, pending, abandoned)

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below:

PROVISIONAL APPLICATION NUMBER	FILING DATE

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorneys, and/or agents with full power of substitution, association, and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected herewith.

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I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application of any patent issued thereon.

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Full Name of Third Joint Inventor, If Any	**Inventor's Signature**	**Date**
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